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Supporting Information

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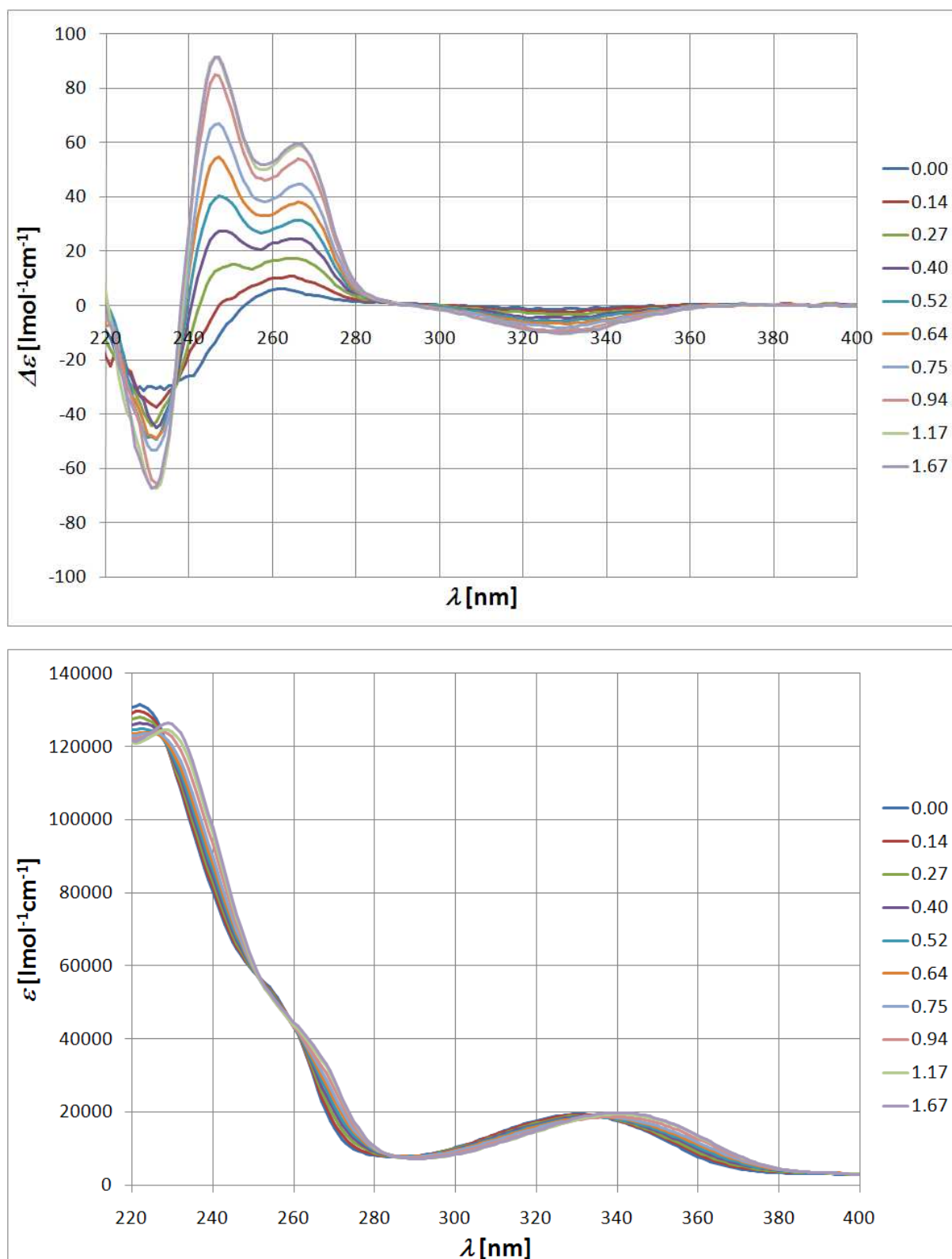
**Synthesis and investigation of a chiral enterobactin analogue based on a  
macrocyclic peptide scaffold**

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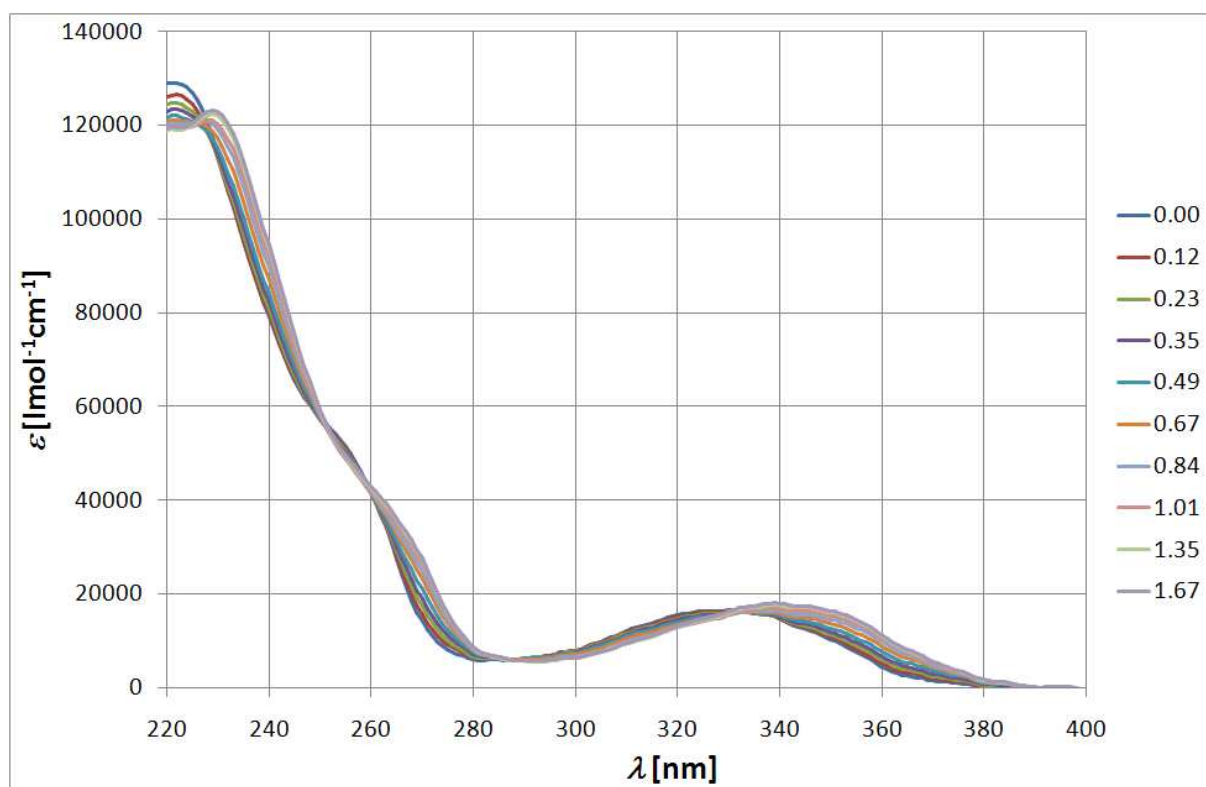
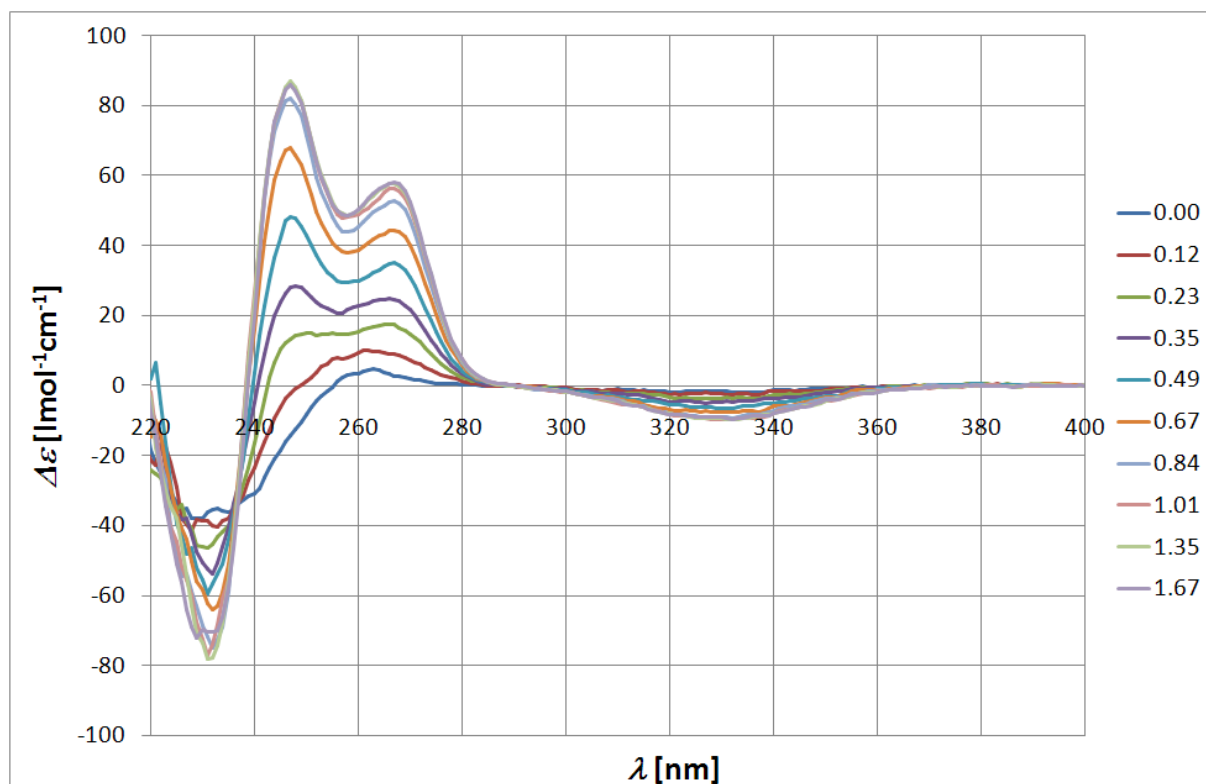
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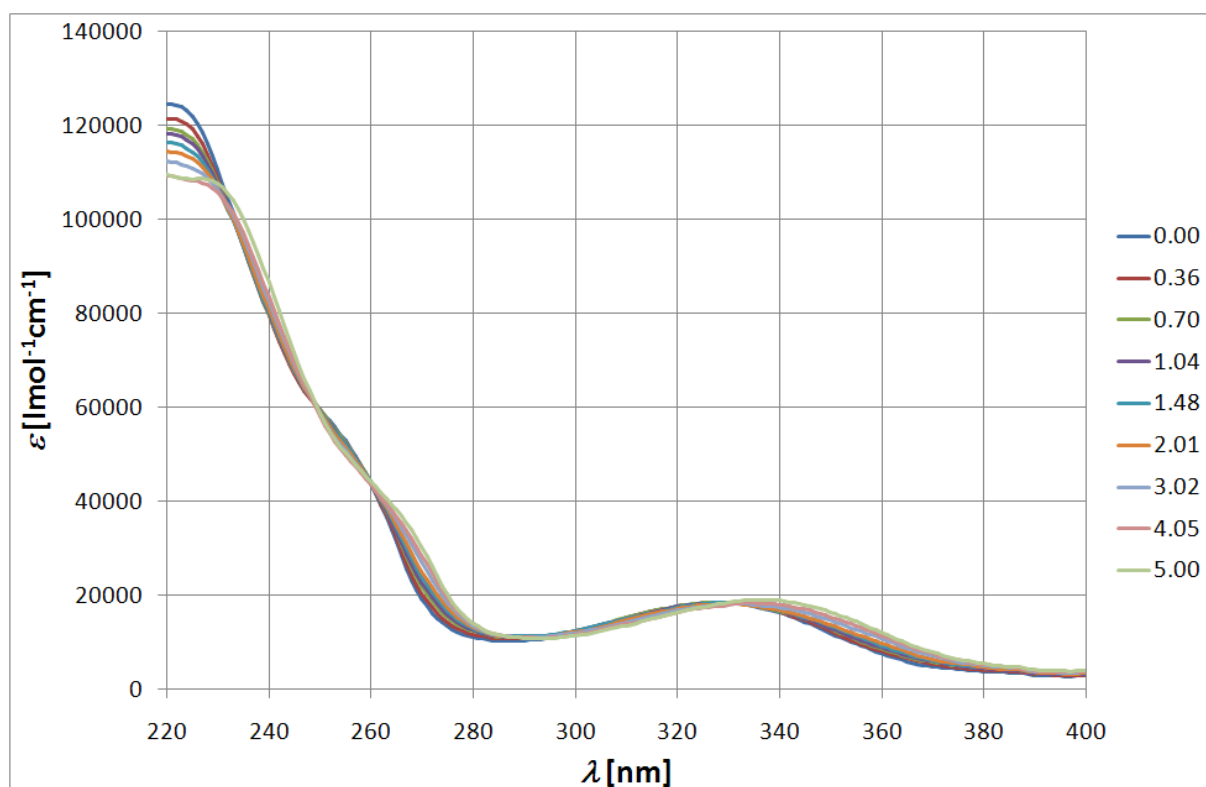
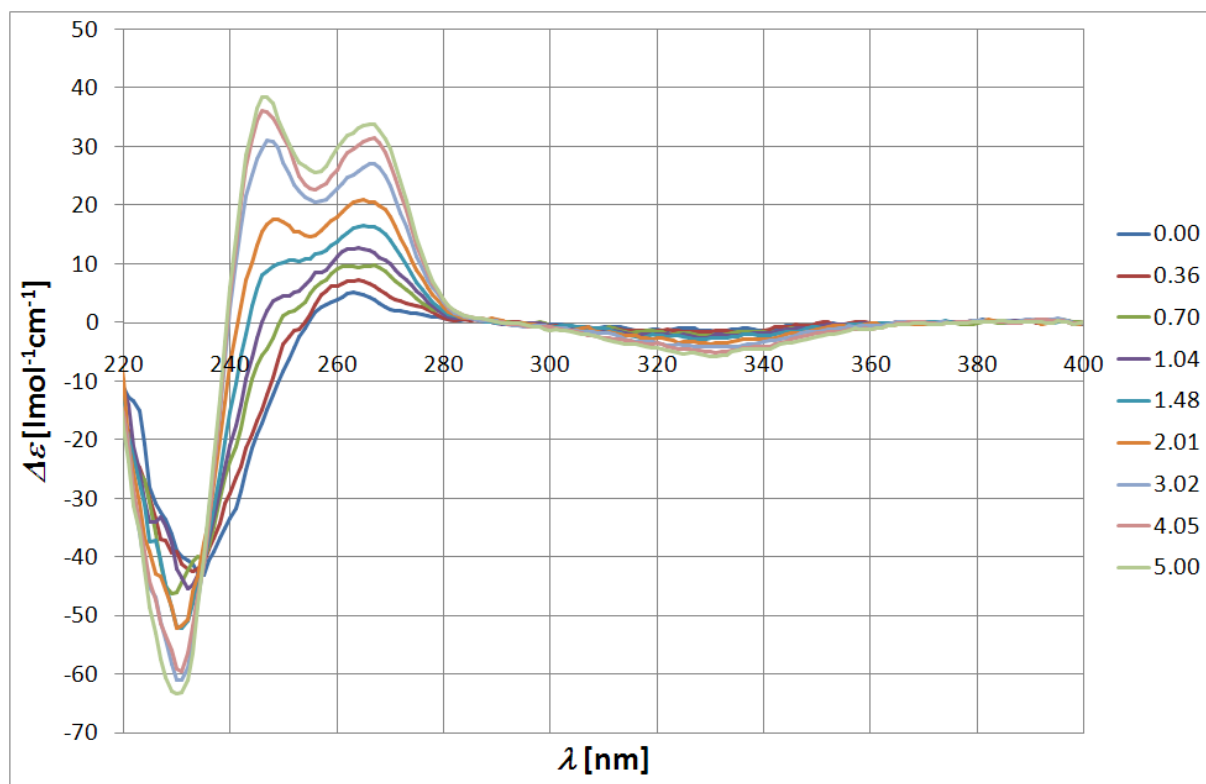
## 1. Spectrophotometric data



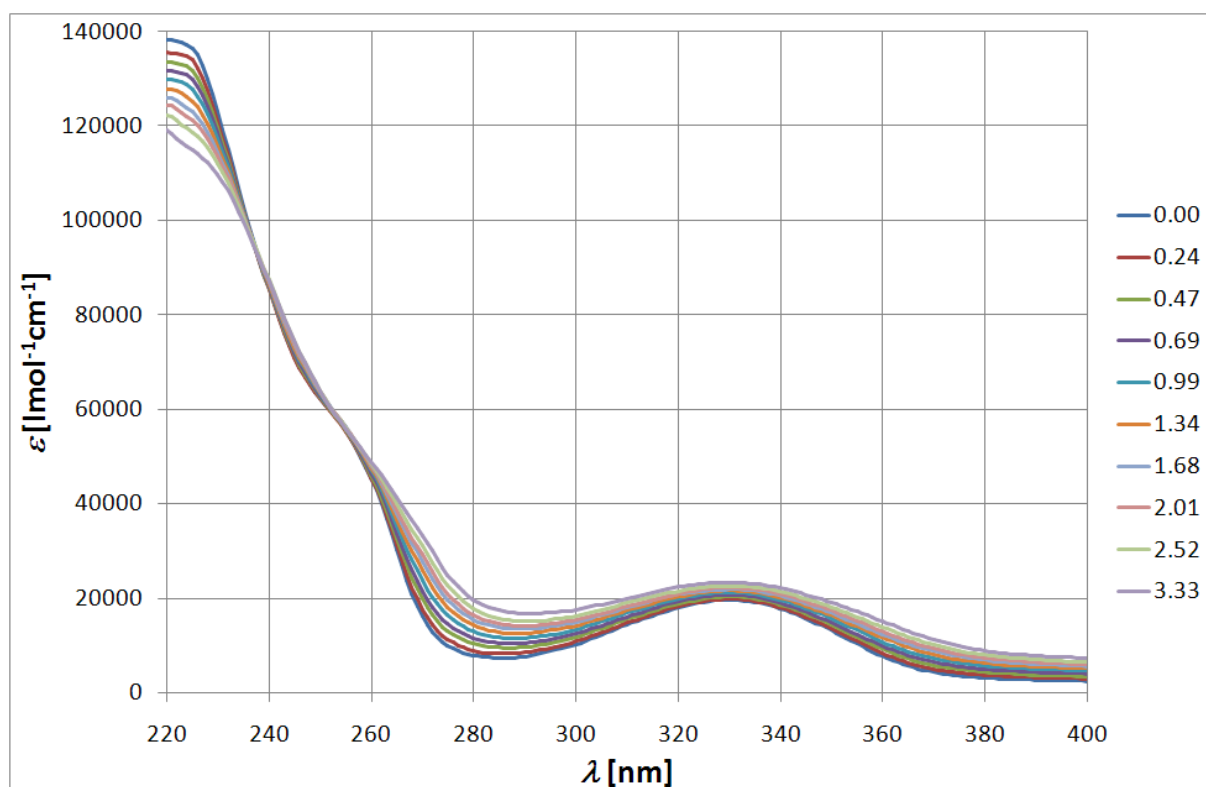
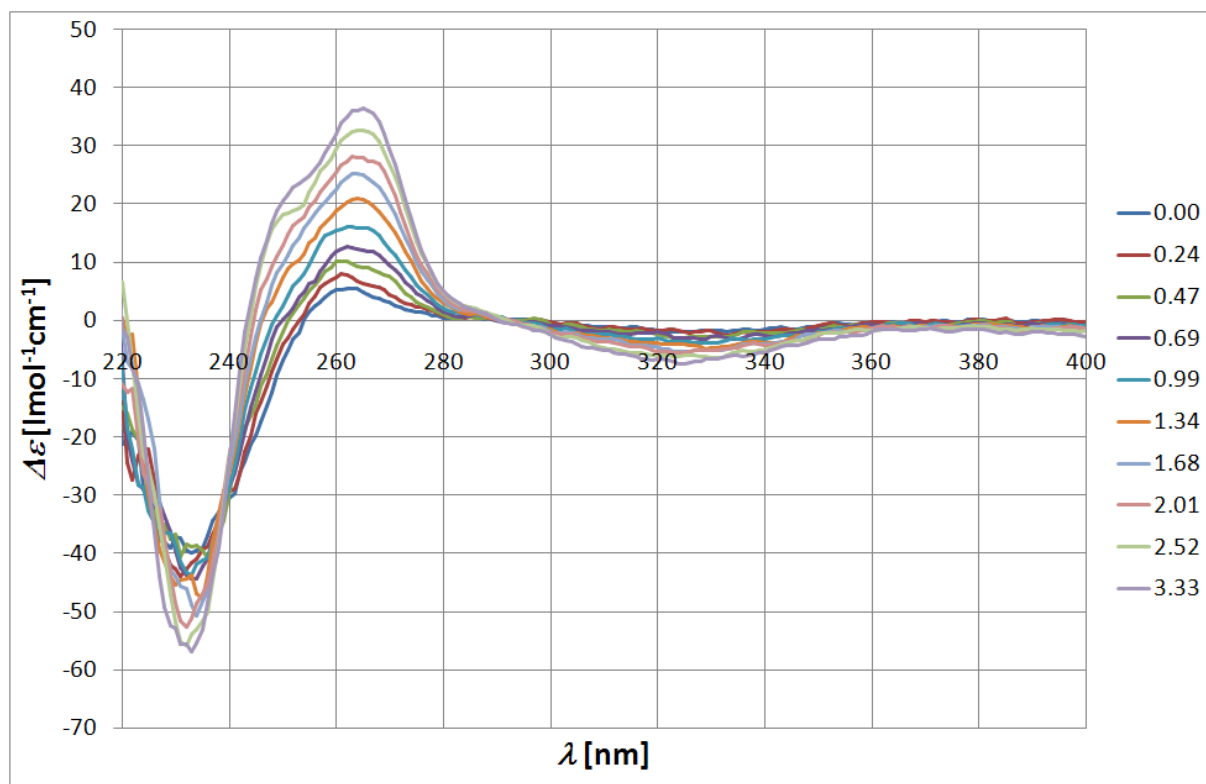
**Figure 1.** Spectrophotometric titration of ligand **1** with  $\text{Al}^{3+}$  ( $[\mathbf{1}] = 2.0 \cdot 10^{-5}$  M, MeOH/ $\text{H}_2\text{O}$  : 10/90; 0.10 M TRIS; 0.02 M HCl buffer; pH = 8.95); top: CD spectra; bottom: UV-vis absorption spectra.



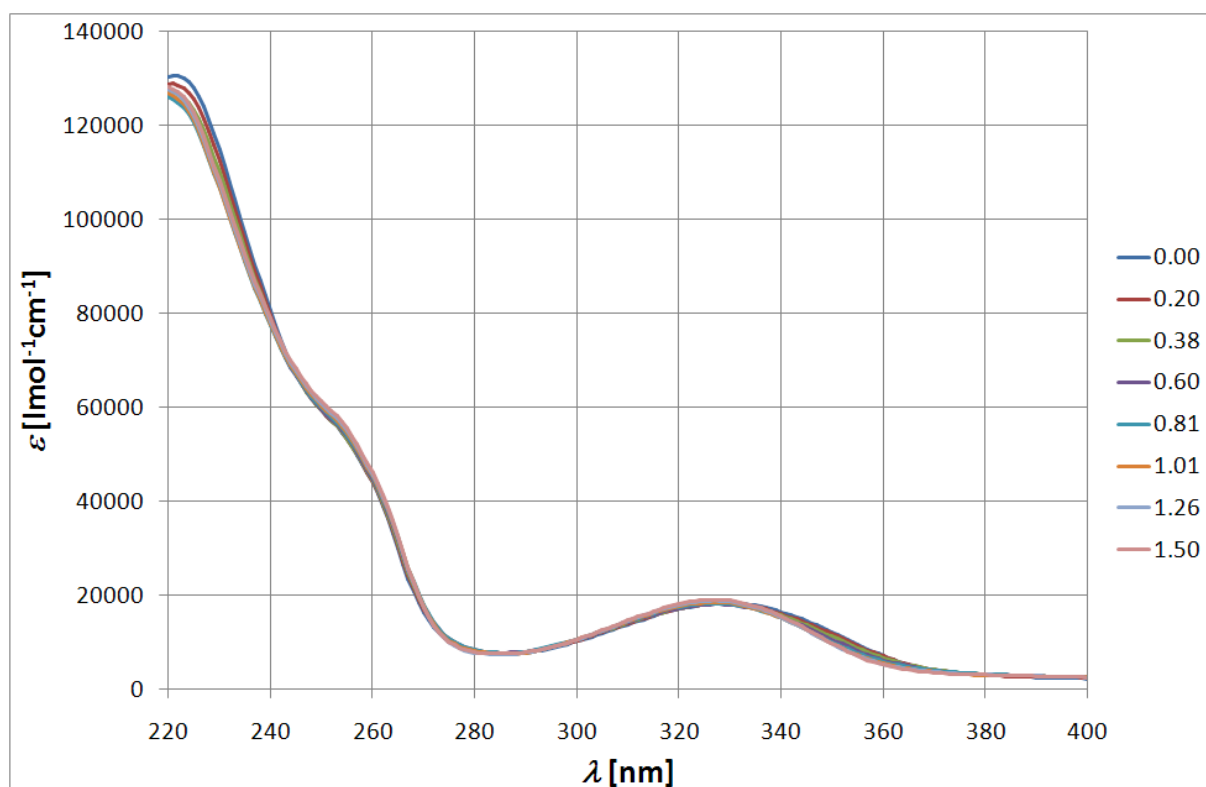
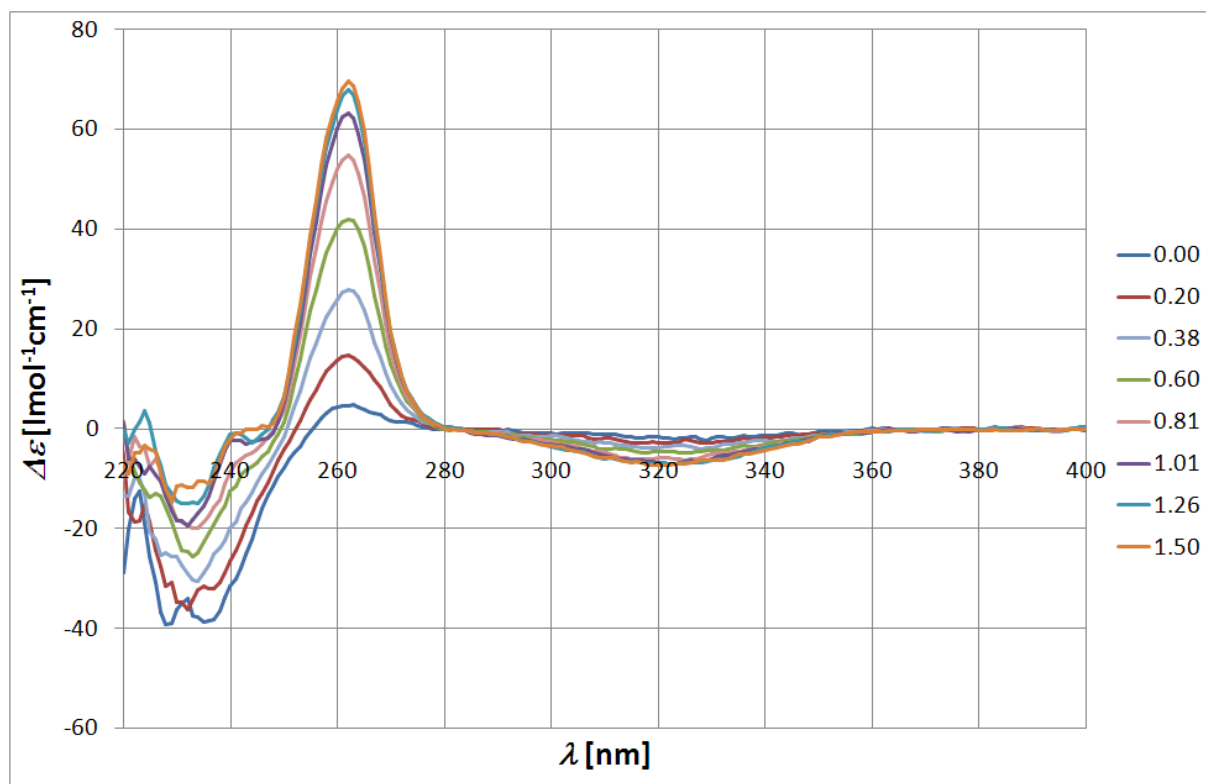
**Figure 2.** Spectrophotometric titration of ligand **1** with  $\text{Ga}^{3+}$  ( $[\mathbf{1}] = 2.0 \cdot 10^{-5} \text{ M}$ , MeOH/ $\text{H}_2\text{O}$  : 10/90; 0.10 M TRIS; 0.02 M HCl buffer; pH = 8.95); top: CD spectra; bottom: UV-vis absorption spectra.



**Figure 3.** Spectrophotometric titration of ligand **1** with  $\text{In}^{3+}$  ( $[\mathbf{1}] = 2.0 \cdot 10^{-5}$  M, MeOH/ $\text{H}_2\text{O}$  : 10/90; 0.10 M TRIS; 0.02 M HCl buffer; pH = 8.95); top: CD spectra; bottom: UV-vis absorption spectra.

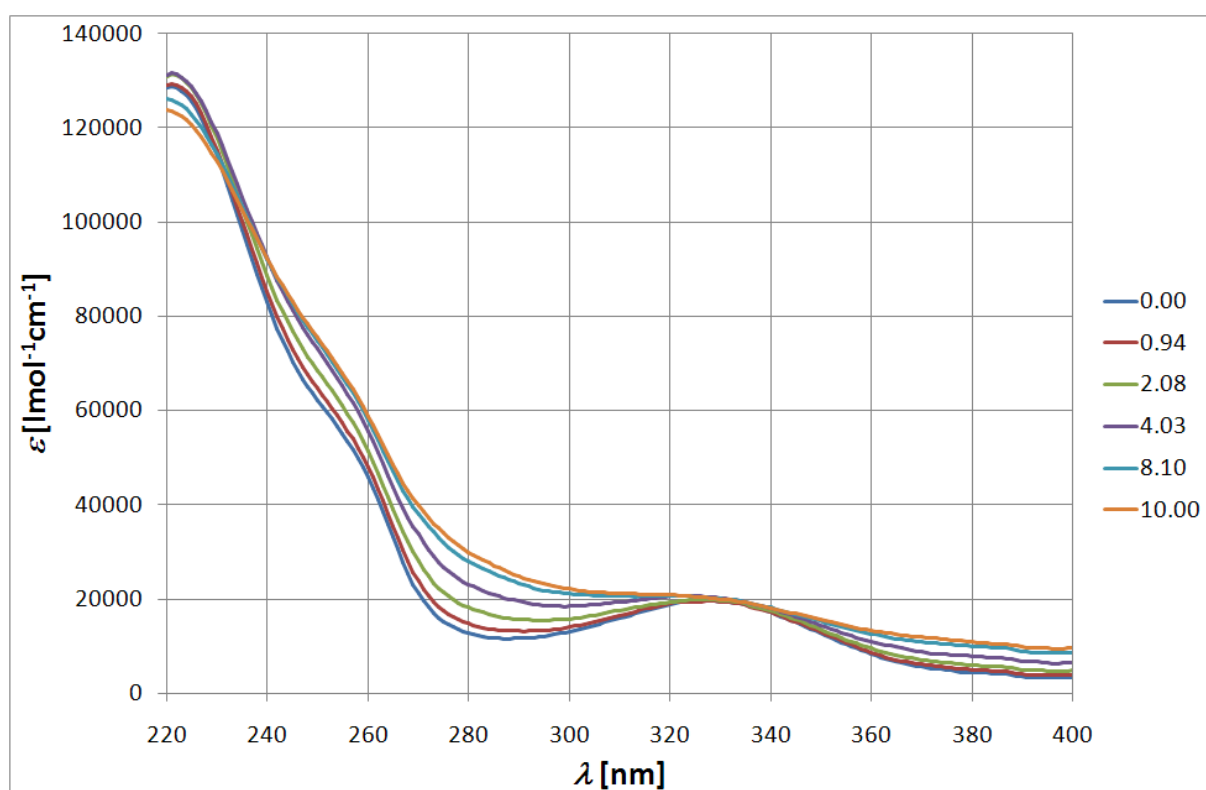
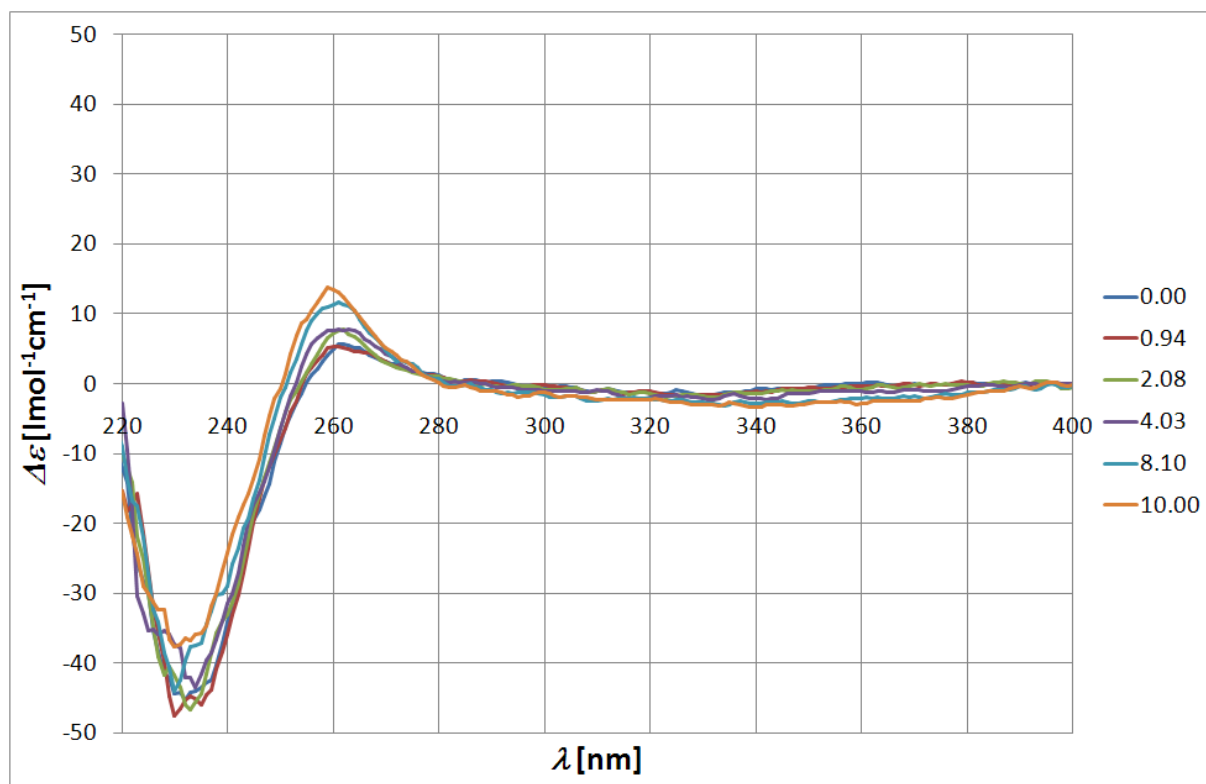


**Figure 4.** Spectrophotometric titration of ligand **1** with  $\text{Fe}^{3+}$  ( $[\mathbf{1}] = 2.0 \cdot 10^{-5}$  M, MeOH/ $\text{H}_2\text{O}$  : 10/90; 0.10 M TRIS; 0.02 M HCl buffer; pH = 8.95); top: CD spectra; bottom: UV-vis absorption spectra.



**Figure 5.** Spectrophotometric titration of ligand **1** with  $\text{Ge}(\text{OCH}_3)_4$  ( $[\mathbf{1}] = 2.0 \cdot 10^{-5}$  M, MeOH/ $\text{H}_2\text{O}$  : 10/90; 0.10 M TRIS; 0.02 M HCl buffer; pH = 8.95); top: CD spectra; bottom: UV-vis absorption spectra.





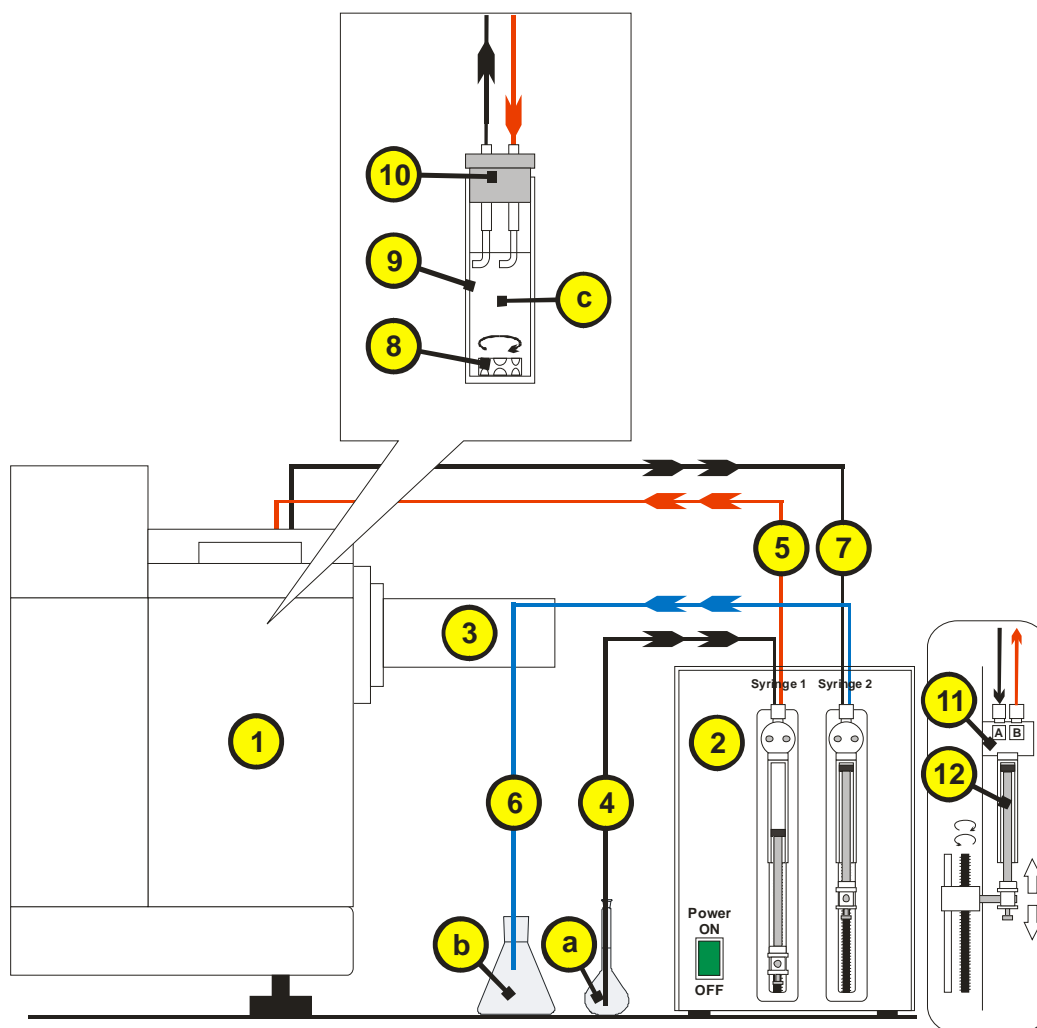
**Figure 6.** Spectrophotometric titration of ligand **1** with  $\text{Ti}(i\text{OPr})_4$  ( $[\mathbf{1}] = 2.0 \cdot 10^{-5} \text{ M}$ ,  $\text{MeOH}/\text{H}_2\text{O} : 10/90$ ,  $0.10 \text{ M TRIS} / 0.02 \text{ M HCl}$  buffer,  $\text{pH} = 8.95$ ); top: CD spectra; bottom: UV-vis absorption spectra.

## 2. Determination of virtual binding constants

Titration experiments for the determination of the association constants were performed with the instrument assembly according to Figure 7. Keeping the concentration of ligand **1** ( $[L] = 2.0 \times 10^{-5} \text{ M}$ ) constant, the metal to ligand molar ratio ( $X = [M]/[L]$ ) was increased from zero to several equivalents in 50 discrete steps. After each addition, molecular spectra were taken after adequate mixing and equilibration time. From the measured UV-vis absorption ( $A(\lambda, X)$ ) and ellipticity ( $\Theta(\lambda, X)$ ) data set, molar extinction ( $\varepsilon(\lambda, X)$ ) and molar circular dichroism ( $\Delta\varepsilon(\lambda, X)$ ) values were calculated according to *Lambert-Beer* law. Equations (1) and (2) expressing explicitly the spectral changes as a function of metal to ligand ratio were used to evaluate the virtual association constants (for a particular wavelength) by nonlinear square fitting to the obtained data  $\Delta\varepsilon^\lambda(X)$  and  $\varepsilon^\lambda(X)$ , respectively. Numeric analysis was performed with Microcal Origin software program by using *Lewenberg-Marquardt* iteration method.

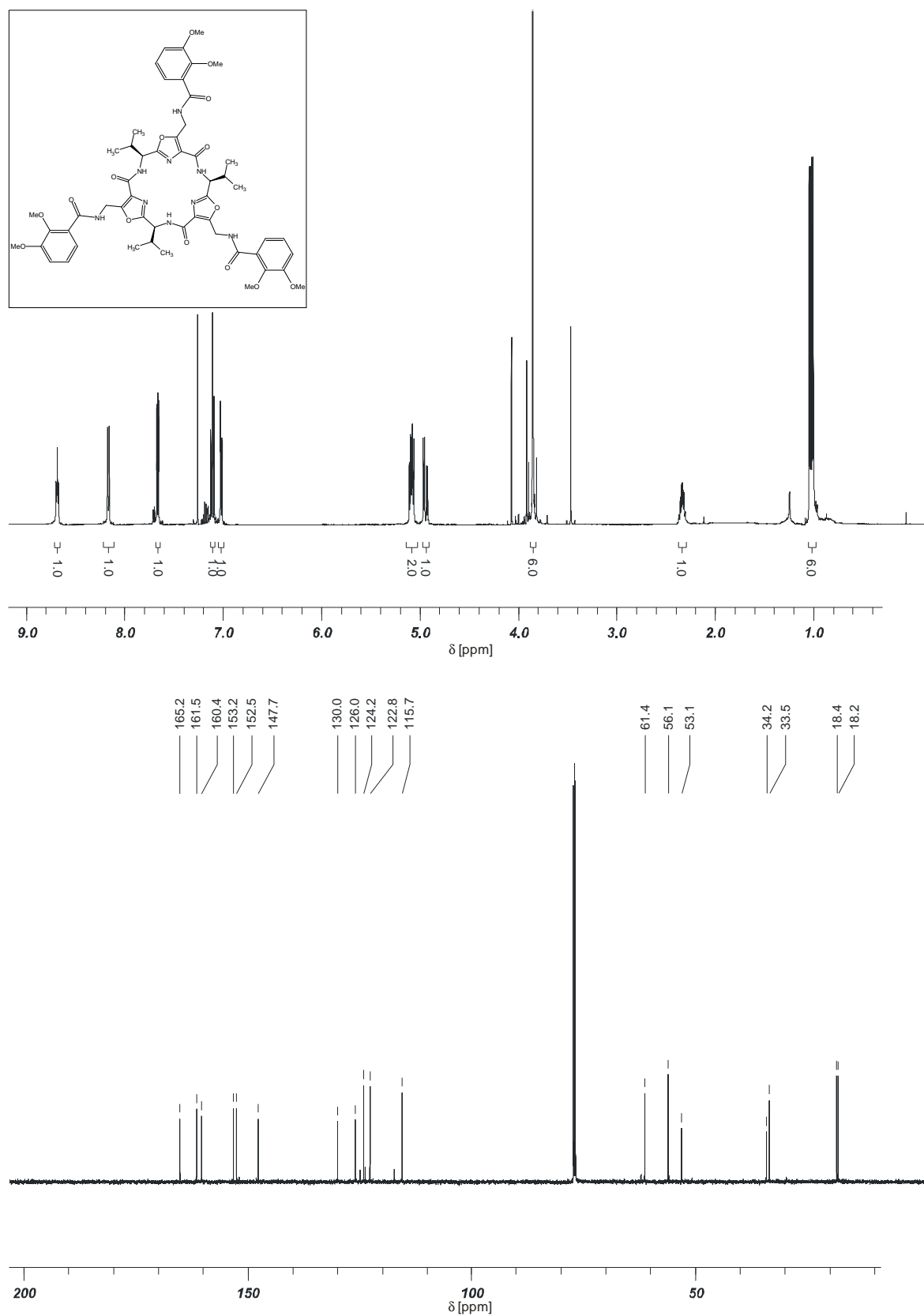
$$\begin{aligned}
 Y^\lambda(X) &= \varepsilon_{obs.}^\lambda(X) = \\
 &= \varepsilon_L^\lambda + \frac{\varepsilon_{ML}^\lambda - \varepsilon_L^\lambda}{2[L]_{tot}} \left\{ \frac{1}{K} + [L]_{tot}(1+X) - \sqrt{\left( \frac{1}{K} + [L]_{tot}(1+X) \right)^2 - 4[L]_{tot}^2 X} \right\} \quad (1)
 \end{aligned}$$

$$\begin{aligned}
 Y^\lambda(X) &= \Delta\varepsilon_{obs.}^\lambda(X) = \\
 &= \Delta\varepsilon_L^\lambda + \frac{\Delta\varepsilon_{ML}^\lambda - \Delta\varepsilon_L^\lambda}{2[L]_{tot}} \left\{ \frac{1}{K} + [L]_{tot}(1+X) - \sqrt{\left( \frac{1}{K} + [L]_{tot}(1+X) \right)^2 - 4[L]_{tot}^2 X} \right\} \quad (2)
 \end{aligned}$$

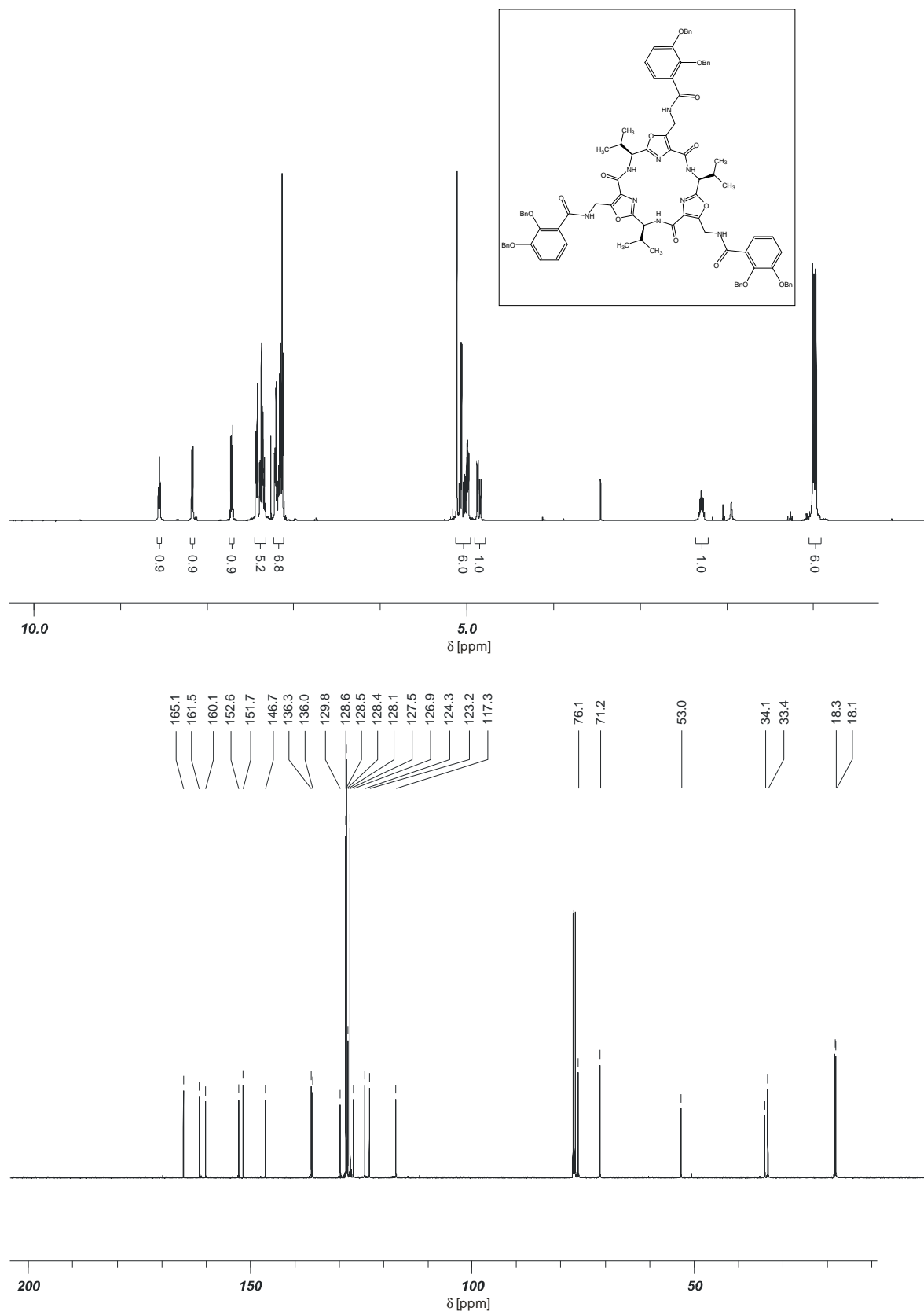


**Figure 7.** 1.) „Jasco J-815“ spectropolarimeter; 2.) „Jasco ATS-443“ automatic titration unit; 3.) detector; 4.) titrant solution intake; 5.) titrant solution inject; 6.) sample solution disposal; 7.) sample solution withdrawal; 8.) stirring magnet; 9.) fluorescence quartz cuvette ( $l = 1 \text{ cm}$ ); 10.) cell cap with nozzles; 11.) magnetic valve (changeover the sample inlet/outlet ports.); 12.) Hamilton Microliter™ syringe ( $V = 2500 \mu\text{l}$ ); a.) titrant solution; b.) waste solution; c.) titrant solution.

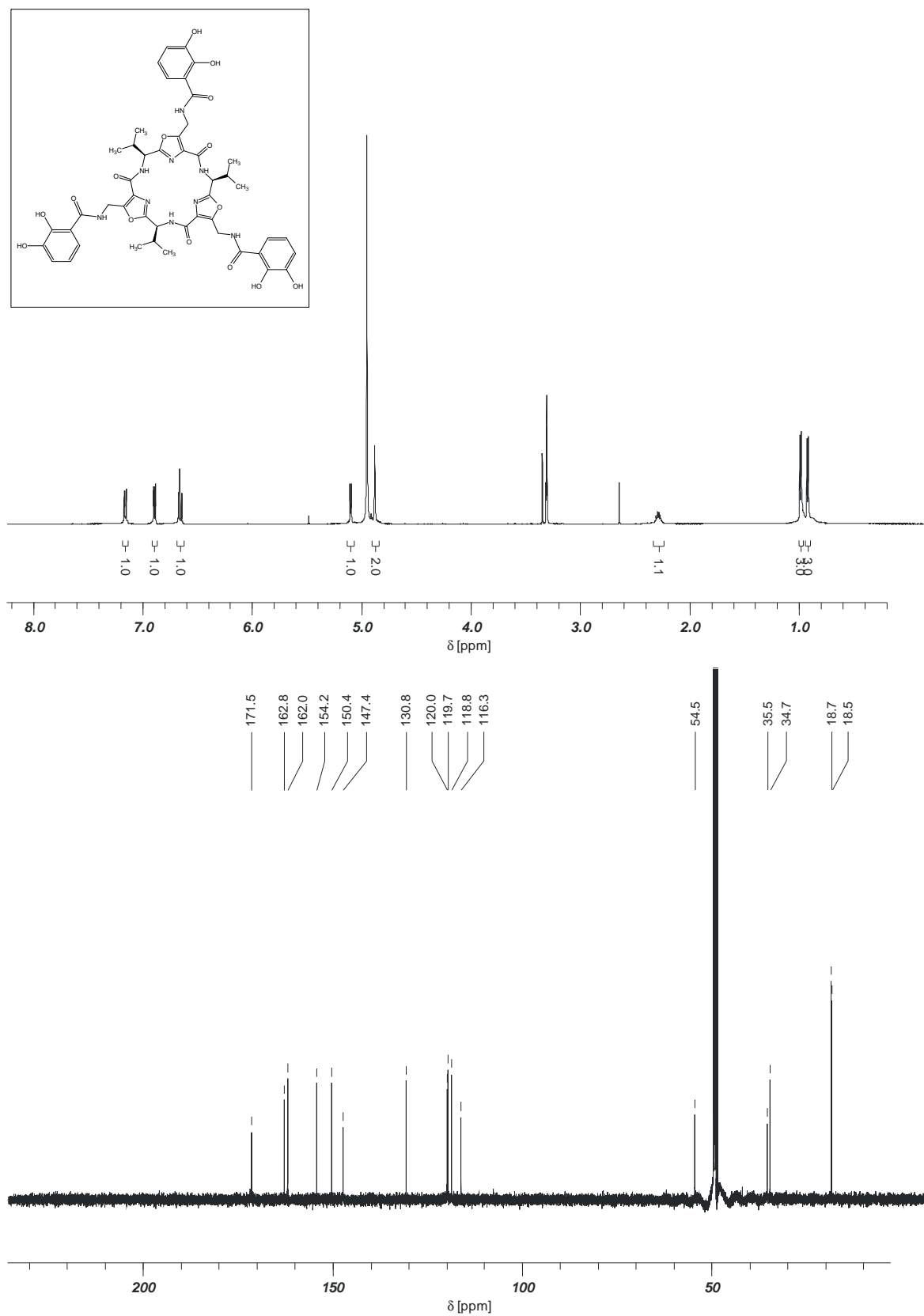
### 3. Nuclear magnetic resonance data



**Figure 8.**  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR spectra of methyl protected ligand **8a** in  $\text{CDCl}_3$ .



**Figure 9.**  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR spectra of benzyl protected ligand **8b** in  $\text{CDCl}_3$ .



**Figure 10.** <sup>1</sup>H- and <sup>13</sup>C-NMR spectra of ligand **1** in MeOH-d<sub>4</sub>.